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Latent microbial infections leading to myelin and axonal damage in multiple sclerosis: A narrative review

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Keywords

Multiple Sclerosis; Therapeutics; Demyelinating Diseases; Axonal Damage; Latent Infections

Abstract

Background: Multiple sclerosis (MS) is a complex autoimmune disease characterized by chronic inflammation, demyelination, and axonal damage in the central nervous system (CNS). This review specifically aims to investigate the role of latent microbial infections-such as those caused by Epstein-Barr virus (EBV), Chlamydia pneumoniae, and others-in contributing to myelin and axon damage in MS.

Methods: We evaluated recent studies from PubMed, Google Scholar, and Scopus databases that focus on the relationship between latent microbial infections and MS pathogenesis.

Results: In MS, emerging evidence suggests that latent microbial infections play a significant role in triggering and perpetuating the inflammatory

processes associated with the disease. The potential mechanisms by which these infections contribute to the pathogenesis of MS, highlighting the interplay between the immune system, microbial agents, and the CNS are evaluated. These include molecular mimicry, where similarities in sequence or structure between viral, bacterial, or self-peptides can activate autoreactive T or B cells through cross activation by pathogen-derived peptides, chronic inflammation triggered by persistent infection, leading to immune-mediated damage, and disruption of the blood-brain barrier, allowing microbial agents or immune cells to infiltrate the CNS.

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Conclusion: This review underscores the critical role of latent microbial infections in MS pathogenesis. By elucidating these mechanisms, we provide new insights that could inform the development of innovative therapeutic interventions and preventive strategies for MS.

Introduction

Background and significance of multiple sclerosis (MS)

MS is a chronic autoimmune disease characterized by inflammation, demyelination, and axonal damage within the central nervous system (CNS). It affects approximately 2.8 million people worldwide, with a higher prevalence in temperate regions, and it predominantly affects young adults.¹ The etiology of MS is multifactorial and remains incompletely understood. However, it is widely accepted that a complex interplay between genetic susceptibility and environmental factors contributes to the development and progression of the disease.²

The pathological hallmark of MS is the destruction of myelin, the protective sheath surrounding nerve fibers, leading to impaired nerve conduction and subsequent axonal degeneration. This process results in the characteristic neurological symptoms observed in MS patients, including weakness, sensory deficits, coordination difficulties, and cognitive impairments.3 The impact of MS goes beyond the physical symptoms, as individuals with the disease often face significant challenges in their daily lives, including decreased quality of life (QOL), increased healthcare utilization, and socioeconomic burden.⁴ Therefore, there is a pressing need to unravel the underlying mechanisms driving MS pathogenesis in order to develop targeted interventions that can halt disease progression, improve symptom management, and enhance overall patient cure.

Emerging evidence suggests that certain viral and bacterial infections may trigger or exacerbate the immune dysregulation observed in MS, leading to an inflammatory cascade that results in autoimmune destruction of myelin and subsequent axonal loss.^{5,6} Understanding the association between latent microbial infections and MS pathogenesis is of utmost importance, as it not only provides insights into the disease mechanisms, but also offers potential therapeutic targets.

Recent breakthroughs in research have highlighted specific microbial agents linked to MS

exacerbations, revealing unresolved questions about their roles in disease pathology. Additionally, controversies surrounding the effectiveness of current diagnostic and therapeutic approaches-particularly regarding their ability to target these latent infections-underscore the necessity for this review.

This narrative review aims to synthesize current findings, highlight gaps in knowledge, and propose directions for future research. By focusing on the relationship between latent microbial infections and MS, we hope to illuminate potential pathways for novel interventions and preventative measures. The information obtained to write this study was collected from reliable databases including Scopus, PubMed, and Google Scholar. **Overview of myelin and axonal damage in MS**

Myelin, a fatty substance produced by specialized cells called oligodendrocytes, forms a protective sheath around nerve fibers in the CNS. It plays a vital role in facilitating rapid and efficient conduction of nerve impulses. In MS, myelin becomes the primary target of autoimmune attack, leading to its destruction and subsequent disruption of nerve signaling.⁷ Activated immune cells, including T cells and macrophages, infiltrate the CNS and release pro-inflammatory cytokines and other mediators, leading to the breakdown of myelin. This process exposes nerve fibers and disrupts their normal functions.⁸

Axonal damage is another critical aspect of MS pathology. Axons, the long projections of nerve cells, transmit electrical signals between different regions of the nervous system. In MS, axonal injury can occur independently or as a consequence of demyelination. The loss of myelin, and the subsequent inflammation, disrupts the normal environment necessary for axonal survival, leading to degeneration and progressive loss of axons.³

Axonal damage in MS is multifactorial and involves various mechanisms. These include direct injury from inflammatory mediators, such as cytokines and free radicals, as well as indirect mechanisms resulting from the loss of trophic support from surrounding myelin. Additionally, immune-mediated mechanisms, such as antibodymediated attacks on axonal proteins, contribute to axonal injury.⁹ The extent of myelin and axonal damage in MS varies among individuals, leading to heterogeneous clinical presentations and disease courses. While early stages of MS may involve remyelination and partial recovery of neurological function, as the disease progresses, chronic inflammation and cumulative damage to myelin and axons result in irreversible disability.¹⁰

Immune-mediated mechanisms of microbialinduced myelin and axonal damage

One of the primary immune-mediated mechanisms involved in microbial-induced myelin and axonal damage is the activation of autoreactive T cells.¹¹ Microbial antigens can activate T cells that recognize both the microbial antigens and self-antigens present in the CNS, including myelin proteins.¹² These activated T cells can infiltrate the CNS, release pro-inflammatory cytokines, and recruit other immune cells, leading to local inflammation and tissue damage.¹¹ Figure 1 illustrates the potential mechanisms by which viruses and bacteria can contribute to the development of MS or trigger exacerbations.

mechanism immune-mediated Another implicated in microbial-induced myelin and axonal production damage is the of autoantibodies. Following microbial infections, B cells can produce antibodies that recognize both microbial antigens and self-antigens present in the CNS. These autoantibodies can bind to myelin and axonal components, leading to complement activation, recruitment of immune cells, and subsequent tissue damage.13 In addition to autoreactive T cells and autoantibodies, other immune cells, such as macrophages and microglia,

play a crucial role in the clearance of microbial pathogens as well as in tissue damage. These immune cells can release pro-inflammatory molecules and cytotoxic factors that contribute to myelin and axonal damage. For example, activated macrophages and microglia can release reactive oxygen species, nitric oxide, and proteases, which can directly damage myelin and axons.14 Moreover, the chronic activation of the immune system in response to persistent or recurrent microbial infections can lead to dysregulation of immune responses. This dysregulation can result in the loss of immune tolerance, leading to the activation of autoreactive immune cells that attack myelin and axonal components in a chronic and sustained manner.15

It is important to note that the specific mechanisms underlying microbial-induced myelin and axonal damage can vary depending on the type of pathogen and the host's genetic and immunological factors. Furthermore, the interplay between microbial infections and other environmental and genetic factors can further modulate the immune responses and contribute to the progression and severity of myelin and axonal damage in neuroinflammatory disorders.¹⁶

The potential mechanisms by which viruses and bacteria can contribute to the development of MS or trigger exacerbations are shown in figure 1.¹⁷

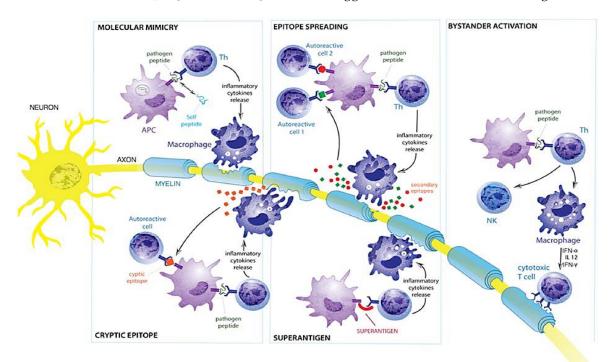


Figure 1. The potential mechanisms by which viruses and bacteria can contribute to the development of MS or trigger exacerbations

One mechanism is molecular mimicry, where similarities in sequence or structure between viral, bacterial, or self-peptides can activate autoreactive T or B cells through cross activation by pathogenderived peptides. Epitope spreading refers to the process of diversifying the immune response from the initial dominant epitope of a protein to secondary epitopes on the same protein (intramolecular spreading) or other proteins (intermolecular spreading). Bystander activation occurs when unrelated infectious agents stimulate natural killer (NK) cells, NKT cells, and macrophages, leading to the production of pro-inflammatory cytokines. These cytokines can non-specifically activate previously primed T or B cells at the target tissue. Subdominant cryptic antigens, usually hidden from the immune system, may be released by antigen-presenting cells (APCs) due to increased protease production in the inflammatory environment caused by infections. Superantigens, which are bacterial, viral, or retroviral proteins, can activate a large proportion of T cells by binding to major histocompatibility complex class II molecules. Unlike classical peptide antigen recognition, superantigens do not require processing into small peptides and can stimulate polyclonal T cell activation and the release of large amounts of cytokines, particularly in T cells expressing specific receptor V β chains. For clarity, the figure does not include other cellular groups involved in the immune response during infections, such as CD8+ cytotoxic T cells, B cells, or microglial cells.

Molecular mimicry and cross-reactivity

Molecular mimicry refers to the structural similarity between microbial antigens and self-antigens, which can result in the activation of autoreactive immune cells that target both the microbial pathogens and components of the CNS.¹⁸ When a microbial infection occurs, the immune system mounts a response to eliminate the invading pathogen. However, in some cases, the immune response can also target self-antigens due to the resemblance between microbial antigens and self-antigens. This occurs when the epitopes of the microbial antigens share molecular similarities, such as amino acid sequences or structural motifs, with self-antigens present in the CNS (Figure 1).¹²

The activation of autoreactive immune cells, particularly T cells, plays a central role in the process of molecular mimicry and cross-reactivity.¹⁸ Autoreactive T cells can be activated by the microbial antigens presented by APCs, and

subsequently, recognize self-antigens in the CNS that share similarity with the microbial antigens. This recognition leads to the activation and expansion of autoreactive T cells, which can infiltrate the CNS and initiate an immune response against myelin and axonal components.^{19,20}

In the context of neuroinflammatory disorders like MS, molecular mimicry and cross-reactivity have been extensively studied. For example, in MS, myelin basic protein (MBP) and myelin oligodendrocyte glycoprotein (MOG) are major target antigens of autoreactive T cells. Several microbial agents, such as human herpesvirus-6 (HHV-6), Epstein-Barr virus (EBV), and certain strains of bacteria, have been implicated in molecular mimicry with MBP and MOG.^{21,22}

Cross-reactivity between microbial antigens and self-antigens can also lead to the activation of autoreactive B cells. B cells can produce autoantibodies that recognize both microbial antigens and self-antigens, including myelin and axonal components. These autoantibodies can contribute to tissue damage by initiating complement activation, opsonization, and antibody-dependent cell-mediated cytotoxicity.²³ The mechanisms underlying molecular mimicry and cross-reactivity involve complex interactions between the immune system, microbial antigens, and self-antigens. Various factors, such as genetic susceptibility, exposure to specific microbial pathogens, and the host's immune response, can influence the development and progression of autoimmune reactions mediated by molecular mimicry.24

Dysregulation of immune responses and chronic inflammation

Dysregulation of immune responses and chronic inflammation play significant roles in the mechanisms underlying microbial-induced myelin and axonal damage in neuroinflammatory disorders. When the immune system fails to effectively control microbial infections or becomes chronically activated, it can lead to sustained inflammation and immune-mediated destruction of myelin and axonal components in the CNS by the continuous activation of immune cells, such as T cells and macrophages.^{11,25} This chronic immune activation results in the release of proinflammatory cvtokines, chemokines. and cytotoxic molecules that contribute to tissue damage. The chronic inflammation can lead to bystander damage of myelin and axons in the CNS, even in the absence of direct infection.26

The dysregulation of immune responses in

microbial-induced myelin and axonal damage involves various mechanisms. One such mechanism is the imbalance between pro-inflammatory and anti-inflammatory signaling pathways. Microbial infections can promote the production of pro-inflammatory cytokines, such as interleukin-1 (IL-1), tumor necrosis factor-alpha (TNF-a), and interferon-gamma (IFN- γ). These cytokines can activate immune cells and amplify the inflammatory response, leading to tissue damage. At the same time, the production of antiinflammatory cytokines, such as IL-10 and transforming growth factor-beta (TGF- β), which normally help regulate inflammation, may be impaired.27-29

Furthermore, dysregulated immune responses can result from defects in immune cell function or regulation. For example, dysfunction of regulatory T cells (Tregs), which play a crucial role in maintaining immune tolerance, can contribute to the breakdown of self-tolerance and the activation of autoreactive immune cells.³⁰ Regulatory T cells help suppress the activation and proliferation of autoreactive T cells, preventing excessive immune responses against self-antigens. In the context of microbial-induced myelin and axonal damage, defects in Treg function can lead to the unchecked activation of autoreactive T cells, perpetuating the inflammatory response and causing tissue damage.³¹

Additionally, dysregulation of the immune response can arise from aberrant activation of complement, a system that plays a role in immune defense and tissue homeostasis.³² In some cases, microbial infections can trigger complement activation, leading to the deposition of complement components on myelin and axonal structures. The activation of complement can initiate an immune response, recruit immune cells, and induce inflammation, which can ultimately contribute to myelin and axonal damage.²¹

The sustained activation of immune cells, such as macrophages and microglia, leads to the release of cytotoxic molecules, including reactive oxygen species, nitric oxide, and proteases. These molecules can directly damage myelin and axonal structures, exacerbating the inflammatory response and further promoting tissue destruction.^{5,11} An important issue is that dysregulation of immune responses and chronic inflammation in the context of microbial-induced myelin and axonal damage can vary depending on the specific microbial pathogen, the host's genetic and environmental influences. factors, The

interplay between these factors contributes to the heterogeneity and complexity of neuroinflammatory disorders.¹⁸

Impact of microbial infections on blood-brain barrier integrity

The integrity of the blood brain barrier (BBB) is crucial for maintaining the homeostasis of the CNS and protecting it from harmful substances.33 Microbial infections can disrupt the BBB, leading to increased permeability and allowing the entry of pathogens, immune cells, and inflammatory mediators into the CNS. This disruption of BBB integrity plays a significant role in the mechanisms underlying microbial-induced myelin and axonal damage neuroinflammatory in disorders.29 Microbial infections can directly affect the BBB through various mechanisms. One mechanism involves the direct invasion of pathogens into the CNS.34 Certain microbial agents including bacteria, viruses, and fungi can breach the BBB by targeting and disrupting the endothelial cells that form the barrier. These pathogens can induce the production of pro-inflammatory cytokines, chemokines, and metalloproteinases matrix (MMPs) that compromise the tight junctions between endothelial cells, leading to increased BBB permeability.35,36

In addition to direct invasion, microbial infections can indirectly impact BBB integrity through the activation of immune cells and the release of inflammatory mediators.³⁷ Following an infection, immune cells, such as monocytes, macrophages, and neutrophils, are recruited to the site of infection. These immune cells can produce inflammatory cytokines, such as IL-1 β and TNF- α , as well as reactive oxygen species and nitric oxide, which can disrupt the tight junctions and compromise BBB integrity.^{38,39}

Furthermore, the activation of immune cells in response to microbial infections can lead to the release of MMPs, enzymes that degrade extracellular matrix components.⁴⁰ MMPs, particularly MMP-2 and MMP-9, can degrade the basement membrane proteins of the BBB, further compromising its integrity and increasing its permeability. This increased permeability allows immune cells and inflammatory mediators to enter the CNS, exacerbating the neuroinflammatory response and contributing to myelin and axonal damage.^{41,42}

The breakdown of BBB integrity also facilitates the entry of microbial toxins into the CNS. Some pathogens produce toxins that directly affect the tight junction proteins and disrupt the BBB.⁴³ For example, certain strains of Escherichia coli produce cytotoxic necrotizing factor (CNF), which can induce the internalization of tight junction proteins and weaken the integrity of the barrier.⁴⁴

The entry of immune cells and inflammatory mediators into the CNS due to BBB disruption leads to the activation of neuroinflammatory processes.⁴⁵ Immune cells, such as T cells and macrophages, can directly target myelin and axonal components, resulting in immune-mediated tissue damage. Additionally, the release of pro-inflammatory cytokines and cytotoxic molecules by immune cells exacerbates the inflammatory response, leading to further myelin and axonal damage.⁵

Latent microbial infections: Unveiling the culprits

Latent microbial infections refer to the presence of microorganisms within the body that can persist in a dormant or inactive state for extended periods. These infections may not cause immediate symptoms or active disease, but can reactivate under certain conditions, leading to pathological consequences.^{46,47} In the context of MS, emerging evidence suggests that latent microbial infections play a significant role in triggering and perpetuating the inflammatory processes associated with the disease.^{17,48,49}

Viruses associated with MS

Latent viral infections have been extensively studied in relation to MS. Several viruses have been implicated, including members of the herpesvirus family. Herpesviruses, such as EBV, HHV-6 human endogenous retrovirus (HERV), varicella-zoster virus, Torque Teno virus (TTV), and cytomegalovirus, have been identified as potential triggers for MS due to their ability to establish lifelong latent infections in the human host.^{17,48,50,51}

EBV is one of the most extensively studied viruses in relation to MS.^{48,52} It is highly prevalent worldwide, with more than 90% of the adult population being infected.⁵³ EBV infection is associated with infectious mononucleosis, and individuals who experience symptomatic primary infection have an increased risk of developing MS later in life.⁵⁴⁻⁵⁶ Furthermore, serological studies have consistently shown a higher prevalence of EBV antibodies in MS patients compared to healthy controls.⁵⁶⁻⁵⁹

The exact mechanisms by which EBV contributes to MS pathogenesis are not fully defined, but several hypotheses have been proposed. One hypothesis suggests that EBV, upon primary infection, may dysregulate the immune system, leading to a cascade of events that ultimately result in an autoimmune response against myelin in susceptible individuals.60 EBV infection has the potential to induce host immune responses, including the production of pro-inflammatory cytokines such as IL-12, IL-1β, IL-6, IL-17, IL-15, GM-CSF, TNF-α, IFN-γ, lymphotoxin-a, and osteopontin through the stimulation of immune-related cell lines. These immune mediators can exacerbate inflammation reactions within MS lesions, leading to neuronal damage and disruption of cellular activity. Furthermore, it is believed that these inflammatory pathways may be activated by sensitization of brain neurons, which could be influenced by genetic alterations associated with MS.17,22,48,61,62

Another hypothesis involves molecular mimicry, where viral antigens resemble self-antigens, leading to cross-reactive immune responses that target both the virus and myelin.57 Autoreactive antibodies in MS can cross-react with viral proteins, particularly EBV nuclear antigen 1 (EBNA1).⁶² Both serum and cerebrospinal fluid (CSF) of MS patients often show elevated levels of antibodies against EBNA1. EBNA1 is consistently recognized as an EBV-specific antigen and stimulates CD4+ T-cell responses in individuals carrying the virus. MS patients exhibit a selective expansion of T cells specific to EBNA1. Additionally, a small subset of these T cells has been found to cross-react with myelin antigens, supporting the hypothesis that clonally expanded EBNA1-specific T cells may play an active role in the immunopathology of MS by promoting cross-recognition through molecular mimicry.63 In addition, during the initial infection, EBV can disrupt the BBB, enabling activated immune cells to enter the CNS, which triggers a series of events that result in CNS inflammation. Moreover, there is evidence to suggest that persistent EBV infection may induce inflammation and immune dysregulation, potentially playing a role in the onset and progression of MS.57 Over-activation of T-cells **B-cells** and during infectious mononucleosis can occur, leading to increased inflammatory reactions in MS.17

HHV-6 is another double-stranded DNA herpesvirus that has been implicated in MS. Like EBV, HHV-6 is highly prevalent and establishes lifelong latency after primary infection. Studies have reported an increased prevalence of HHV-6 DNA in the blood and CSF of MS patients compared to controls.⁶⁴ HHV-6 infection has been associated with increased disease activity and exacerbations in MS, suggesting a potential role in disease pathogenesis.65

The mechanisms by which HHV-6 influences MS are not abundantly understood, but may involve direct viral-induced damage, activation of immune responses, or interactions with other viral or environmental factors.^{66,67} HHV-6 has been shown to infect and replicate within astrocytes, microglia, and oligodendrocytes, leading to cellular damage and inflammation.⁶⁸ According to study findings, the continuous presence of active HHV-6 infection in glial cells within inflamed CNS tissue could potentially lead to virus-induced immune-pathologies in MS.⁶³

It is worth noting that a remarkable similarity exists in the amino acid sequence of HHV-6 24-hour urine (U24) protein and MBP, which is considered a potential autoantigen associated with MS. Moreover, there is evidence of cross-reactivity between autoreactive T-cells and MBP, indicating the possibility of a molecular mimicry mechanism in HHV-6 infection.⁶⁹ HHV-6 infection may induce the production of pro-inflammatory cytokines and chemokines, contributing to the perpetuation of the inflammatory processes observed in MS.⁶⁵

In the context of HHV-6 infection, virusinfected T lymphocytes exhibit an upregulation of proinflammatory genes, including IL-1, IL-2, IL-18, IFN, and TNF-a, while simultaneously downregulating anti-inflammatory cytokines such as IL-10 and IL-14. The elevated production of these inflammatory mediators by immune cells is responsible for triggering intense inflammatory reactions, leading to the development of demyelination and damage to nerve myelin and axons.61 There is much evidence indicating a correlation between levels of inflammatory cytokines, including TNF-a, in the CSF and the degree of disability and progression rate observed in patients with MS.66 The response of CD8+ T lymphocytes to HHV-6-infected CNS cells can lead to tissue injury and the release of sequestered antigens. This, in turn, activates self-reactive lymphocytes and enhances autoreactive immune reactions. Activation of the complement system can be improved through the utilization of CD46, which is used by HHV-6A as a cellular receptor.52 Viruses involved in MS are presented in table 1 in terms of virus type, involvement with the disease, and relationship with MS.

Bacterial infections and their role in MS pathogenesis Bacterial infections have also been implicated in the development and progression of MS. Chronic infections, such as those caused by Chlamydia pneumoniae, Mycoplasma pneumoniae, Helicobacter pylori, Borrelia burgdorferi, Mycobacterium tuberculosis (TB), and Mycobacterium avium have been shown to be associated with MS and have been detected in MS patients. These bacteria can persist within cells and tissues, evading the immune system and potentially contribute to chronic inflammation and tissue damage.^{17,48,49,70-72}

The exact mechanisms by which bacterial infections contribute to MS pathogenesis are not fully understood. However, several hypotheses have been proposed. One hypothesis suggests that bacterial infections may trigger an immune response that leads to chronic inflammation. This chronic inflammation can subsequently damage myelin and contribute to the development of MS.⁷³ Another hypothesis involves molecular mimicry, where bacterial antigens resemble self-antigens, leading to cross-reactive immune responses that target both the bacteria and myelin.⁷⁴

Chlamydia pneumoniae is an intracellular bacterium that can cause respiratory tract infections. This bacterium has been detected in the CNS of MS patients, suggesting its potential involvement in the disease.¹⁷ Studies have shown an increased prevalence of C. pneumoniae antibodies in MS patients compared to healthy individuals.⁴⁸ Furthermore, experimental models have demonstrated that C. pneumoniae infection can induce a persistent infection in the brain, and consequently, induce immune responses and promote CNS inflammation, which may contribute to pathogenesis of chronic inflammatory diseases such as MS.^{36,75}

Chlamydia pneumoniae has the ability to enhance the expression of MHC class II molecules CD40, CD80, and CD86 on bone marrow-derived dendritic cells (BMDDC). This, in turn, induces the production of proinflammatory cytokines by macrophages. The secretion of these cytokines can increment the frequency of T-lymphocytes, leading to the recognition of various antigens, including antigens that react with self-proteins. This process has the potential to trigger autoimmune disorders.⁷⁶

Chlamydia pneumoniae has the ability to infect various types of cells, including macrophages, monocytes, endothelial cells, and smooth muscle cells in blood vessels. As a result, infected monocytes/macrophages, responding to an initial trigger such as an acute viral infection or autoimmune reaction, could transport this pathogen to inflamed CNS tissues.⁷⁷

trom Bar-Or et al. ⁶⁰)		
Virus	Disease involvement	Association with MS
EBV, HHV-4, lymphocryptovirus Double-stranded (ds) DNA virus,	Infectious mononucleosis, Hodgkin and non-Hodgkin's	Mononucleosis predisposes to MS, reduced activity of EBV-specific
neurotropic Tropism: B cells,	lymphoma, Burkett's	cytotoxic T cells (e.g., exhausted) in
epithelial cells	lymphoma, gastric and	MS patients, EBV seropositive
Cell latency: memory B cells	nasopharyngeal cancer, hairy	epidemiological studies, virus present
Centratency. memory B cens	cell leukemia, MS	in MS brain, EBV prolongs the lifespan of B cells
HHV-6, roseolovirus	Exanthema subitum	Present in MS plaques, reactivation
dsDNA, neurotropic Tropism:	(roseola infantum) and	during relapses, high levels found in
broad; hematopoietic and	pneumonitis, MS	oligodendrocytes and areas of
epithelial cells	-	demyelination, elevated levels are
Cell latency: lymphocytes		found early in MS and during
and monocytes		relapses/exacerbations, anti-HPV IgG
		and IgM titers are reported to predict
		relapses
CMV, betaherpesvirinae	Retinitis, hepatitis, colitis,	Both detrimental and beneficial
dsDNA, neurotropic Tropism: broad;	pneumonia, encephalitis, MS	properties reported, large meta-analysis
hematopoietic cells, smooth muscle,		MS versus controls did not yield a
monocytes, epithelial and endothelial		conclusive link between CMV and MS
cells, fibroblasts, connective tissue		
Cell latency: cells of the myeloid lineage		
VZV, HHV-3	Chickenpox, shingles, MS	Virus is present during relapses, recent
dsDNA, neurotropic Tropism:		studies failed to show an increased risk
mononuclear cells		of MS associated with varicella or
Cell latency: sensory ganglia		zoster infections
HERV-W	MS, diabetes, autoimmune	Present in infiltrating macrophages and
Tropism: cells of the nervous system,	arthritis, and schizophrenia	activated MS lesions, MSRV Env
syncytiotrophoblast layer of the placenta	In most cases the observed	protein is detected in blood of active
Cell latency: multiple	expression profiles of specific	MS patients, drives the expression of
	HERV-W sequences have not	proinflammatory cytokines, reduces
	led to a definitive association	myelin protein, expression and kills
	with human disease	oligodendrocyte precursors
	pathology.	

Table 1. Viruses implicated in MS: Virus, disease involvement, and association with multiple sclerosis (MS) (Adapted from Bar-Or et al.⁶⁰)

MS: Multiple sclerosis; EBV: Epstein-Barr virus; VZV: Varicella zoster virus; HHV-3: Human herpesvirus-3; CMV: Cytomegalovirus; IgM: Immunoglobulin M; HPV: Human papillomavirus; HERV: Human endogenous retrovirus

Recent reports have shown that C. pneumoniae can infect glial cells and ependymal cells within the CNS of mice.⁷⁰ The bacterium was noted to cross the BBB and enter the CNS, disrupting its permeability by releasing toxic components from its cell wall.49 This process leads to the increased production of inflammatory factors by activated microglia, which can have either neurotoxic or neuroprotective effects depending on the disease stage.⁷⁸ The presence of C. pneumoniae in the CNS can initiate an initial inflammatory response and also serve as a chronic stimulus, leading to a sustained state of immune activation. On the other hand, CNS infection by C. pneumoniae in patients with MS may simply be a secondary infection of already damaged CNS tissue.77

Mycoplasma pneumonia is a small

microorganism that adheres to host cells by specific attachments and adhesin antigens that has been associated with MS.⁴⁹ This bacterium can cause respiratory tract infections and has been detected in the CNS of MS patients.^{76,79} Studies have reported an increased prevalence of M. pneumoniae antibodies in MS patients compared to controls.¹⁷ Mycoplasma infections have been shown to induce immune responses and promote inflammation, potentially contributing to the development or exacerbation of MS.⁸⁰

The lipoproteins of M. pneumoniae play a significant role in infection and modulation of immunity through TLR1 and TLR2. The co-expression of TLR2 and TLR6 mediates the cellular response to lipopeptides from M. pneumoniae, which have been recognized in

cerebral endothelial cells and microglia of MS patients.⁴⁹ Additionally, M. pneumoniae has been found to invade the CNS and is known to induce demyelination, at least in the peripheral areas. It is considered a possible co-factor in the development of MS.⁸¹

Helicobacter pylori is a gram-negative bacterium that resides on the surface of gastric epithelial cells and is generally considered a bacterium, non-invasive but laboratory observations have shown that it can enter the epithelial and immune cells of the host.82 While a number of studies have not found an association between H. pylori and susceptibility to MS, several reports have demonstrated a high incidence of acute H. pylori infection and significantly higher frequency of H. pylori immunoglobulin G (IgG) seropositivity in remitting-relapsing and secondary progressive MS patients during the stable phase, compared to healthy people.49,83-85 Some studies indicate that H. pylori infection leads to a decrease in the levels of Th1 and Th17 cells in the CNS and spleen, in comparison to the control group. Consequently, these findings have prompted suggestions of a potential protective effect of H. pylori against MS.17,86 Persistent bacterial infection can lead to a loss of self-tolerance due to the continuous release of bacterial antigens capable of motivating the release of pro-inflammatory cytokines from immune cells. H. pylori can apply these effects not only locally, but also directly on the CNS, modulating the brain-gut axis.87

Borrelia burgdorferi is the bacteria responsible for Lyme disease, a syndrome that shares similarities with MS. The bacterium activates macrophages through TLR2 and stimulates Th1-type T-lymphocytes immunity. During an infection, B. burgdorferi employs various mechanisms to manipulate the innate and adaptive immune systems, enabling its survival within mammalian host cells.48,88,89 In some patients, Borrelia infection can trigger an autoimmune response and the release of inflammatory mediators, leading to the improvement of chronic neurological abnormalities similar to MS. This is according to the observation that both Lyme disease and MS are related to abnormal immune reactions, and some individuals with MS have stated a history of Lyme disease or exposure to disease vectors. Both disorders exhibit myelin damage and inflammation, which can make it challenging to differentiate between Lyme

borreliosis and MS.⁴⁸ Several studies have indicated an increase in anti-Borrelial antibody levels in MS patients, suggesting a potential relationship between Lyme disease and MS.^{48,90}

It is important to consider Lyme disease during the differential diagnosis of MS, as it mimics several neurological symptoms. However, the presence of antibodies against B. burgdorferi in MS patients does not confirm current or past infections, nor does it prove that the bacteria is the cause of the disease.⁴⁹

Mycobacterium TB is an exceptionally successful pathogen that can persist in host tissues for years without triggering disease. This bacterium is responsible for TB, an infectious and contagious disease primarily affecting the lungs.⁹¹ TB is categorized into two types: latent tuberculosis infection (LTBI) and active TB disease. LTBI refers to a persistent immune response to M. tuberculosis antigens lacking clinically manifested active TB disease.⁹²

The connection between TB and MS can be understood in two aspects. Firstly, immunomodulation or immunosuppression treatments may lead to the reactivation of latent TB infection. Secondly, the intense inflammatory response of the body prior to bacillus infection can increase susceptibility to autoimmune diseases like MS. This occurs because there is a similarity observed between epitopes from the chaperone HSP60 of M. tuberculosis and fragments of HSP60 found in MS patients. Notably, a specific peptide of the bacillus binds strongly to multiple alleles, suggesting its contribution in the pathogenesis of MS by inducing a robust immune response.71 Several studies have demonstrated an association between mycobacteria and MS. Components of mycobacteria have the ability to activate the innate immune system through toll-like receptors (TLRs). Stimulation of the host immune response via TLR2, TLR4 and TLR9 trigger cytokines production, leading to the differentiation of naive CD4+ T cells into Th1 and Th17 cells. Consequently, this results in the production of IL-17 and IFN-y. These combined factors facilitate the migration of leukocytes across the BBB, contributing to the activation of inflammatory immune responses, tissue damage and neuronal dysfunction in MS disease.93,94 Regarding their possible role in MS, several studies have shown a higher frequency of lymphocyte proliferative response against recombinant proteins HSP70 and HSP65 derived from M. tuberculosis in MS patients compared to those with other neurological disorders or healthy individuals.^{93,95}

Mycobacterium avium subsp. Paratuberculosis (MAP), classified as an intracellular pathogen within the M. avium complex, is a non-tuberculous mycobacterium responsible for paratuberculosis in ruminants. Additionally, it has been suggested as the potential cause of Crohn's disease and other chronic inflammatory infections in humans.96 Recent research indicates that MAP may have a causative role in MS pathology, particularly in genetically susceptible individuals, according to the theory of molecular mimicry. Currently, clinical trials are underway to investigate antimycobacterial therapy targeting MAP.93 The involvement of mycobacteria in the development and progression of MS may be specific to certain populations and strongly influenced by various genetic and non-genetic factors. However, the potential of immune modulation as an approach to combat mycobacterial infection remains largely unexplored.93

Furthermore, MAP peptides have been found to elicit a T cell response in peripheral blood mononuclear cells (PBMCs) isolated from individuals with relapsing-remitting MS.97 Additionally, MS patients have shown the presence of intrathecally synthesized IgG antibodies that react to specific MAP-derived peptides.98 In the context of MAP infection or antigen exposure, autoreactive T-cells activated in the peripheral immune system through molecular mimicry may cross the BBB and become reactivated in the CNS by local APCs.49

Regarding the invasion of MAP, it primarily occurs through M cells located in the Peyer's patches. Subsequently, it invades intra-epithelial macrophages where it can proliferate. The recognition of MAP involves TLR2 and NOD2, which bind to mannosylated-lipoarabinomannan (Man-LAM) and peptidoglycans, respectively.^{49,99} Moreover, MAP has the aptitude to inhibit MyD88 and TLR9 signaling and modulate the expression of IFN- γ receptors, permitting evasion of both innate and acquired immune responses in cattle.⁴⁹

The specific roles of various TLRs and MyD88mediated immune responses in MS, as well as the impact of MAP antigens on experimental autoimmune encephalomyelitis (EAE), are still subjects of investigation. One theory proposes that chronic MAP infection in the human gastrointestinal tract may induce the release of proinflammatory mediators, leading to neurons inflammation and polarization of the immune response toward a Th1/Th17 phenotype.¹⁰⁰

It is important to note that while there is evidence supporting the association between bacterial infections and MS, causality has not been definitively established. The presence of bacterial DNA or antibodies in MS patients may reflect a response to infection or colonization rather than direct causation. Further research is needed to better understand the relationship between bacterial infections and MS.

Other potential microbial triggers: fungi and parasites

In addition to viruses and bacteria, other microbial agents, such as fungi and parasites, have been investigated for their potential involvement in MS pathogenesis. Although the evidence is limited, studies have reported associations between fungal infections, such as those caused by Candida species, Aspergillus and Cryptococcus neoformans.^{101,102} Similarly, parasitic infections, such as those caused by helminths, have investigated been for their potential immunomodulatory effects and their influence on MS development and progression.^{103,104}

Candida species are opportunistic pathogens that can cause superficial and systemic infections, while C. neoformans is a fungal pathogen associated with meningitis. Studies have shown an increased prevalence of fungal infections and elevated levels of anti-fungal antibodies in MS patients compared to healthy controls.¹⁰⁵ Antibodies against Candida designate a past fungal infection which can generate memory B-lymphocyte diagnosing epitopes of fungal infection. Memory B-cells which recognize both a fungal epitope and an epitope in the nerve cells might elucidate the association between Candida antibodies and MS disease.106

Parasitic infections have also been explored as potential triggers of immune dysregulation in MS. Parasites such as Toxoplasma gondii and Plasmodium spp. (the causative agents of toxoplasmosis and malaria, respectively) have been studied in relation to MS. Toxoplasma gondii infection has been associated with an increased risk of developing MS and exacerbation of disease activity, although the results of several studies suggest а negative relationship among Toxoplasma gondii infection and MS which can be considered as a likely protective feature of toxoplasmosis against the improvement of MS disease.107-110 Plasmodium infection has been suggested to have a protective effect against MS development due to its ability to induce immunomodulatory responses.^{111,112} These findings highlight the complex interactions between parasites and the immune system in the context of MS. Therefore, some microorganisms, such as the mentioned parasites, can play a disease suppressor role against MS. More extensive studies are needed to achieve reliable results and confirm the association between these microbes and MS disease.

The mechanisms by which fungi and parasites may contribute to MS pathogenesis are still being investigated. It is hypothesized that these microorganisms can trigger immune responses that result in chronic inflammation and immune dysregulation. Additionally, molecular mimicry and cross-reactivity between microbial antigens and self-antigens may lead to autoimmune reactions targeting both the microorganism and myelin components.¹¹³

Diagnostic methods for identifying latent infections in MS

Serological and molecular techniques: Various serological and molecular techniques have been employed to investigate the presence and potential contribution of latent infections in MS. These diagnostic methods provide valuable insights into the association between infectious agents and the or progression of MS.^{114,115} development Serological techniques involve the detection of specific antibodies in blood samples to determine exposure or immune response to particular infectious agents. In the context of latent infections in MS, serological testing can help identify the presence of antibodies against specific pathogens. For instance, the presence of antibodies against common viruses such as EBV or HHV-6 may indicate prior exposure or reactivation of these viruses in individuals with MS. Enzyme-linked immunosorbent assay (ELISA), immunofluorescence assays, and Western blotting are commonly used serological techniques to detect specific antibodies.116,117

Molecular techniques play a crucial role in identifying the presence of latent infections by detecting the genetic material or specific markers of infectious agents. Polymerase chain reaction (PCR) is a widely used molecular technique that amplifies and detects specific DNA or RNA sequences. In the context of MS and latent infections, PCR can be utilized to detect viral DNA or RNA in CSF or PBMCs. For example, PCR-based assays can detect the presence of viral genetic material, such as EBV or HHV-6 DNA, in biological samples from individuals with MS. Real-time PCR, nested PCR, and quantitative PCR (qPCR) are variations of PCR commonly employed in diagnostic settings.^{115,118,119}

In addition to PCR, other molecular techniques such as next-generation sequencing (NGS) have emerged as powerful tools in the identification of latent infections. NGS allows for the comprehensive analysis of the entire microbial community within a sample, enabling the detection of known and potentially novel infectious agents. By sequencing the genetic material present in a sample, NGS can provide a more comprehensive view of the microbial landscape associated with MS. This technique has the potential to identify previously unrecognized infectious agents and elucidate their potential role in MS pathogenesis.120

It is important to note that the identification of latent infections in MS using serological and molecular techniques has its challenges and limitations. Serological assays may have limitations in terms of sensitivity and specificity, as the presence of antibodies does not necessarily indicate active infection or a causal relationship with MS. Molecular techniques, while highly sensitive, may also suffer from false positives and negatives, depending on factors such as the timing of sample collection, handling, and assay design. Additionally, the presence of viral genetic material alone does not establish causality, as viral reactivation or persistence may occur without direct involvement in MS pathology.

Imaging modalities for detecting CNS infections

In addition to serological and molecular techniques, imaging modalities play a crucial role in the identification and evaluation of latent infections in MS. These imaging techniques allow for the visualization of CNS structures and the detection of abnormalities associated with infectious processes. Magnetic resonance imaging (MRI) is the primary imaging modality used in the diagnosis and monitoring of MS. MRI can also provide valuable insights into CNS infections associated with MS. Various MRI sequences and contrast agents can be applied to detect specific findings indicative infectious imaging of processes. These findings may include the presence of parenchymal lesions, periventricular or juxtacortical white matter abnormalities, or enhancement patterns suggestive of active inflammation.121 Contrast-enhanced MRI with

gadolinium is particularly useful in detecting areas of BBB disruption and assessing the extent of inflammatory processes.

In the context of latent infections in MS, MRI can help identify specific characteristics associated with CNS infections. For example, certain infectious agents, such as progressive multifocal leukoencephalopathy (PML) caused by JC virus, can exhibit characteristic imaging features on MRI. PML typically presents as asymmetrical white matter lesions with a predilection for the subcortical and periventricular regions.¹²² These lesions often lack mass effect or significant enhancement, which can help differentiate them from other MS-related lesions.

In addition to conventional MRI, advanced imaging techniques can provide further insights into the pathophysiology of CNS infections. Diffusion-weighted imaging (DWI) and diffusion tensor imaging (DTI) can assess the microstructural integrity of brain tissue and detect changes associated with infectious processes. Quantitative measures derived from DWI and DTI, such as apparent diffusion coefficient (ADC) and fractional anisotropy (FA), respectively, can provide valuable information about tissue damage and inflammation.123 Functional MRI (fMRI) techniques can assess alterations in cerebral perfusion and neuronal activity, aiding in the characterization of infection-related changes in brain function.

Positron emission tomography (PET) is another imaging modality that can contribute to the evaluation of CNS infections in MS. PET scans, combined with specific radiopharmaceuticals, can detect metabolic changes associated with infectious processes. For instance, fluorodeoxyglucose (FDG) PET can assess glucose metabolism and identify areas of increased metabolic activity, which may be indicative of infectious or inflammatory foci.¹²⁴ Other specific radiotracers, such as [11C]-PK11195, can target microglial activation and provide insights into the inflammatory response associated with CNS infections.

It is important to note that while imaging modalities can provide valuable information about CNS infections in MS, they are not definitive diagnostic tools on their own. The interpretation of imaging findings should be considered in conjunction with clinical history, serological and molecular test results, and other diagnostic data. Furthermore, imaging findings may not always be specific to a particular infectious agent and can overlap with MS-related abnormalities.

Therapeutic approaches targeting latent infections in MS

Antiviral therapies: Antiviral medications aim to suppress viral replication and reduce viralinduced inflammation. For example, ganciclovir and valganciclovir have been explored for the treatment of HHV-6-associated MS, while antiviral drugs like acyclovir and valacyclovir have been investigated for their potential benefits in EBV-associated MS. These medications may help control viral activity and potentially modify disease course. Immunomodulatory therapies, such as IFN-beta (IFN- β) and glatiramer acetate, which are commonly used in MS treatment, have been suggested to have antiviral effects. These medications may exert their therapeutic effects by modulating the immune response against viral infections and suppressing viral replication. By reducing viral-induced inflammation, these immunomodulatory therapies may indirectly impact the progression of MS. A combination of antiviral medications targeting HHV-6 with immunomodulatory therapies such as IFN- β has been investigated in clinical trials to assess their efficacy in controlling viral infections and modifying the course of MS.^{125,126}

Another important aspect of therapeutic approaches targeting latent infections in MS is personalized medicine. As MS is a heterogeneous disease with variations in viral associations, disease progression, and treatment response, individualized treatment strategies may be necessary. By identifying specific viral infections and their impact on an individual's disease, personalized therapy can be tailored to target existing specific latent infection(s). This may involve selecting the most appropriate antiviral medication or combination therapy based on the viral profile and disease characteristics of each patient.¹²⁷

Antibiotics and immunomodulators

In addition to antiviral therapies, other therapeutic approaches have been explored to target latent infections in MS. These approaches of antibiotics involve the use and immunomodulators, which aim to modulate the immune response, reduce inflammation, and potentially alleviate MS symptoms.128,129 Tetracyclines, such minocycline as and doxycycline, have been investigated for their potential therapeutic effects in MS. These antibiotics have antimicrobial both and immunomodulatory properties. They can suppress bacterial infections associated with MS, such as C. pneumoniae, and also modulate the immune response and reduce inflammation. Tetracyclines have been shown to inhibit immune cell activation and migration, reduce the production of proinflammatory cytokines, and promote antiinflammatory responses. Clinical trials assessing the efficacy of tetracyclines in MS are ongoing.¹³⁰

Macrolide antibiotics, including azithromycin and clarithromycin, have also been studied in the context of MS. Like tetracyclines, macrolides possess antimicrobial and immunomodulatory properties. They can target bacterial infections associated with MS, such as M. pneumoniae and C. pneumoniae and modulate the immune response. Macrolides have been shown to inhibit the production of pro-inflammatory cytokines, reduce immune cell activation, and promote regulatory T-cell responses. Clinical trials investigating the use of macrolides in MS are currently underway.¹³¹

IFN-β is a widely used immunomodulatory therapy in MS. It can reduce inflammation, modulate the immune response, and potentially impact latent infections. IFN-β has been shown to have antiviral properties by inhibiting viral replication and promoting an antiviral state within cells. Additionally, it can modulate the immune response against viral infections. Clinical studies have suggested that IFN-β treatment may reduce viral activity and potentially modify the course of MS in individuals with latent infections.¹³²

checkpoint Immune inhibitors, such as natalizumab and fingolimod, are immunomodulatory therapies that target specific molecules involved in immune cell migration and activation. These medications have been investigated for their potential effects on latent infections in MS. By modulating immune cell trafficking and reducing immune cell infiltration into the CNS, these therapies may indirectly impact viral replication and inflammation associated with latent infections. However, caution must be exercised when using immune checkpoint inhibitors, as they can increase the risk of certain viral infections.133

Combination therapies and future directions

Combination therapies and future directions in the treatment of latent infections in MS have gained attention as potential strategies to target the underlying viral involvement and modify the disease course. By combining different therapeutic approaches, such as antiviral therapies, immunomodulators, and other emerging treatments, it is hoped that a synergistic effect can be achieved to control viral replication, modulate the immune response, and potentially alleviate MS symptoms.¹³⁴

Personalized medicine holds great promise in the treatment of MS and latent infections. By identifying specific viral infections and their impact on an individual's disease, tailored treatment strategies can be developed. This may involve selecting the most appropriate combination therapy based on the viral profile, disease characteristics, and treatment response of each patient. Personalized medicine can optimize treatment efficacy and minimize any potential adverse effects.¹³⁵

Another future direction is the investigation of immune tolerance induction strategies. These approaches aim to reprogram the immune system to tolerate viral infections and reduce the inflammatory response in MS. Techniques such as antigen-specific immunotherapy, regulatory T-cell induction, and immune modulation through microbiota may potentially be effective in promoting immune tolerance and mitigate viralassociated inflammation in MS.¹³⁶

Conclusion

In this narrative review, we highlight the growing evidence supporting the involvement of latent microbial infections in the pathogenesis of MS, specifically focusing on myelin and axonal damage observed in this disease. By elucidating the underlying mechanisms and exploring potential diagnostic methods and therapeutic interventions, we aimed to contribute to a better understanding of MS and pave the way for improved management strategies that target latent infections. Further research is warranted to unravel the complex interactions between microbial agents and the immune system in MS, ultimately, leading to improved patient outcomes and the development of novel treatment approaches.

Conflict of Interests

The authors declare no conflict of interest in this study.

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References

- Piggott T, Nonino F, Baldin E, Filippini G, Rijke N, Schunemann H, et al. Multiple Sclerosis International Federation guideline methodology for off-label treatments for multiple sclerosis. Mult Scler J Exp Transl Clin 2021; 7(4): 20552173211051855.
- Goncalves JNC, Cortez P, Carvalho MS, Fraz+úo NM. A multivariate approach for multi-step demand forecasting in assembly industries: Empirical evidence from an automotive supply chain. Decision Support Systems 2021; 142: 113452.
- Nicholson TR, Carson A, Edwards MJ, Goldstein LH, Hallett M, Mildon B, et al. Outcome measures for functional neurological disorder: A review of the theoretical complexities. J Neuropsychiatry Clin Neurosci 2020; 32(1): 33-42.
- Lakin L, Davis BE, Binns CC, Currie KM, Rensel MR. Comprehensive approach to management of multiple sclerosis: addressing invisible symptoms-a narrative review. Neurol Ther 2021; 10(1): 75-98.
- Correale J, Marrodan M, Ysrraelit MC. Mechanisms of neurodegeneration and axonal dysfunction in progressive multiple sclerosis. Biomedicines 2019; 7(1): 14.
- Milo R, Korczyn AD, Manouchehri N, Stuve O. The temporal and causal relationship between inflammation and neurodegeneration in multiple sclerosis. Mult Scler 2020; 26(8): 876-86.
- Franklin RJ, Ffrench-Constant C. Remyelination in the CNS: from biology to therapy. Nat Rev Neurosci 2008; 9(11): 839-55.
- Lassmann H. Pathogenic mechanisms associated with different clinical courses of multiple sclerosis. Front Immunol 2018; 9: 3116.
- Frischer JM, Bramow S, Dal-Bianco A, Lucchinetti CF, Rauschka H, Schmidbauer M, et al. The relation between inflammation and neurodegeneration in multiple sclerosis brains. Brain 2009; 132(Pt 5): 1175-89.
- Hauser SL, Oksenberg JR. The neurobiology of multiple sclerosis: Genes, inflammation, and neurodegeneration. Neuron 2006; 52(1): 61-76.
- Wekerle H, Lassmann H. The immunology of inflammatory demyelinating disease. In: Compston A, Confavreux C, Lassmann H, McDonald I, Miller D, Noseworthy J, et al., editors. McAlpine's Multiple Sclerosis. 4th ed.Edinburgh, UK: Churchill Livingstone; 2006. p. 491-555.
- Venigalla SSK, Premakumar S, Janakiraman V. A possible role for autoimmunity through molecular mimicry in alphavirus mediated arthritis. Sci Rep 2020; 10(1): 938.
- Lucchinetti CF, Mandler RN, McGavern D, Bruck W, Gleich G, Ransohoff RM, et al. A role for humoral mechanisms in the pathogenesis of Devic's neuromyelitis

optica. Brain 2002; 125(Pt 7): 1450-61.

- Hoftberger R, Lassmann H, Berger T, Reindl M. Pathogenic autoantibodies in multiple sclerosis - from a simple idea to a complex concept. Nat Rev Neurol 2022; 18(11): 681-8.
- 15. Kim JS, Soto-Diaz K, Bingham TW, Steelman AJ, Das A. Role of omega-3 endocannabinoids in the modulation of T-cell activity in a multiple sclerosis experimental autoimmune encephalomyelitis (EAE) model. J Biol Chem 2023; 299(2): 102886.
- 16. Tiwari S, Lapierre J, Ojha CR, Martins K, Parira T, Dutta RK, et al. Signaling pathways and therapeutic perspectives related to environmental factors associated with multiple sclerosis. J Neurosci Res 2018; 96(12): 1831-46.
- Marrodan M, Alessandro L, Farez MF, Correale J. The role of infections in multiple sclerosis. Mult Scler 2019; 25(7): 891-901.
- Sundaresan B, Shirafkan F, Ripperger K, Rattay K. The Role of Viral Infections in the Onset of Autoimmune Diseases. Viruses 2023; 15(3): 782.
- Moudgil KD. Viewing autoimmune pathogenesis from the perspective of antigen processing and determinant hierarchy. Crit Rev Immunol 2020; 40(4): 329-39.
- Goverman J. Autoimmune T cell responses in the central nervous system. Nat Rev Immunol 2009; 9(6): 393-407.
- Tarlinton RE, Martynova E, Rizvanov AA, Khaiboullina S, Verma S. Role of viruses in the pathogenesis of multiple sclerosis. Viruses 2020; 12(6): 643.
- Soldan SS, Lieberman PM. Epstein-Barr virus and multiple sclerosis. Nat Rev Microbiol 2023; 21(1): 51-64.
- 23. Maciak K, Pietrasik S, Dziedzic A, Redlicka J, Saluk-Bijak J, Bijak M, et al. Th17-related cytokines as potential discriminatory markers between neuromyelitis optica (Devic's Disease) and multiple sclerosis-a review. Int J Mol Sci 2021; 22(16): 8946.
- Getts DR, Spiteri A, King NJC, Miller SD. Chapter 21 - microbial infection as a trigger of T-Cell Autoimmunity. In: Rose NR, Mackay IR, editors. The autoimmune diseases. 6th ed. Academic Press; 2020. p. 363-74.
- Venkatesan A, Johnson RT. Infections and multiple sclerosis. Handb Clin Neurol 2014; 122: 151-71.
- Mangale V, McIntyre LL, Walsh CM, Loring JF, Lane TE. Promoting remyelination through cell transplantation therapies in a model of viral-induced neurodegenerative disease. Dev Dyn 2019; 248(1): 43-52.
- Dinarello CA. Proinflammatory cytokines. Chest 2000; 118(2): 503-8.
- Michalickova D, Sima M, Slanar O. New insights in the mechanisms of impaired redox signaling and its interplay with inflammation and immunity in multiple

sclerosis. Physiol Res 2020; 69(1): 1-19.

- van Langelaar LJ, Rijvers L, Smolders J, van Luijn MM. B and T cells driving multiple sclerosis: identity, mechanisms and potential triggers. Front Immunol 2020; 11: 760.
- Schloder J, Shahneh F, Schneider FJ, Wieschendorf B. Boosting regulatory T cell function for the treatment of autoimmune diseases - That's only half the battle! Front Immunol 2022; 13: 973813.
- 31. Verma ND, Lam AD, Chiu C, Tran GT, Hall BM, Hodgkinson SJ. Multiple sclerosis patients have reduced resting and increased activated CD4(+) CD25(+) FOXP3(+)T regulatory cells. Sci Rep 2021; 11(1): 10476.
- Saez-Calveras N, Stuve O. The role of the complement system in Multiple Sclerosis: A review. Front Immunol 2022; 13: 970486.
- Segarra M, Aburto MR, Acker-Palmer A. Blood-brain barrier dynamics to maintain brain homeostasis. Trends Neurosci 2021; 44(5): 393-405.
- 34. Tran VTA, Lee LP, Cho H. Neuroinflammation in neurodegeneration via microbial infections. Front Immunol 2022; 13: 907804.
- Cai Z, Qiao PF, Wan CQ, Cai M, Zhou NK, Li Q. Role of blood-brain barrier in Alzheimer's Disease. J Alzheimers Dis 2018; 63(4): 1223-34.
- 36. Dando SJ, Mackay-Sim A, Norton R, Currie BJ, St John JA, Ekberg JA, et al. Pathogens penetrating the central nervous system: infection pathways and the cellular and molecular mechanisms of invasion. Clin Microbiol Rev 2014; 27(4): 691-726.
- 37. da Fonseca AC, Matias D, Garcia C, Amaral R, Geraldo LH, Freitas C, et al. The impact of microglial activation on blood-brain barrier in brain diseases. Front Cell Neurosci 2014; 8: 362.
- 38. Liu Q, Gao Y, Zhang B, Sun F, Yang Q, Liu Y, et al. Cytokine profiles in cerebrospinal fluid of patients with meningitis at a tertiary general hospital in China. J Microbiol Immunol Infect 2020; 53(2): 216-24.
- Gerhauser I, Hansmann F, Ciurkiewicz M, Loscher W, Beineke A. Facets of Theiler's murine encephalomyelitis virus-induced diseases: an update. Int J Mol Sci 2019; 20(2): 448.
- Muri L, Leppert D, Grandgirard D, Leib SL. MMPs and ADAMs in neurological infectious diseases and multiple sclerosis. Cell Mol Life Sci 2019; 76(16): 3097-116.
- 41. Krueger M, Mages B, Hobusch C, Michalski D. Endothelial edema precedes blood-brain barrier breakdown in early time points after experimental focal cerebral ischemia. Acta Neuropathol Commun 2019; 7(1): 17.
- Alvarez JI, Katayama T, Prat A. Glial influence on the blood brain barrier. Glia 2013; 61(12): 1939-58.
- 43. Gay F. Bacterial toxins and Multiple

Sclerosis. J Neurol Sci 2007; 262(1-2): 105-12.

- Chaoprasid P, Dersch P. The Cytotoxic Necrotizing Factors (CNFs)-A Family of Rho GTPase-Activating Bacterial Exotoxins. Toxins (Basel) 2021; 13(12).
- 45. Takata F, Nakagawa S, Matsumoto J, Dohgu S. Blood-brain barrier dysfunction amplifies the development of neuroinflammation: Understanding of Cellular events in brain microvascular endothelial cells for prevention and treatment of BBB dysfunction. Front Cell Neurosci 2021; 15: 661838.
- Roe K. A latent pathogen infection classification system that would significantly increase healthcare safety. Immunol Res 2023; 71(5): 673-7.
- Khabibullina NF, Kutuzova DM, Burmistrova IA, Lyadova IV. The biological and clinical aspects of a latent tuberculosis infection. Trop Med Infect Dis 2022; 7(3): 48.
- Landry RL, Embers ME. The probable infectious origin of multiple sclerosis. NeuroSci 2023; 4(3): 211-34.
- Cossu D, Yokoyama K, Hattori N. Bacteria-host interactions in multiple sclerosis. Front Microbiol 2018; 9: 2966.
- Bjornevik K, Munz C, Cohen JI, Ascherio A. Epstein-Barr virus as a leading cause of multiple sclerosis: mechanisms and implications. Nat Rev Neurol 2023; 19(3): 160-71.
- Kakalacheva K, Munz C, Lunemann JD. Viral triggers of multiple sclerosis. Biochim Biophys Acta 2011; 1812(2): 132-40.
- Rosella M, Carmela R, Roberta R, Grazia M, Maria CB, Silvia R, et al. Viruses and neuroinflammation in multiple sclerosis. Neuroimmunology and Neuroinflammation 2021; 8: -269.
- 53. Shi T, Huang L, Tian J. Prevalence of Epstein-Barr Viral DNA among children at a single hospital in Suzhou, China. J Pediatr (Rio J) 2022; 98(2): 142-6.
- Ascherio A, Munger KL. Epstein-Barr virus infection and multiple sclerosis: A review. J Neuroimmune Pharmacol 2010; 5(3): 271-7.
- 55. Guan Y, Jakimovski D, Ramanathan M, Weinstock-Guttman B, Zivadinov R. The role of Epstein-Barr virus in multiple sclerosis: from molecular pathophysiology to in vivo imaging. Neural Regen Res 2019; 14(3): 373-86.
- 56. Thomas OG, Rickinson A, Palendira U. Epstein-Barr virus and multiple sclerosis: moving from questions of association to questions of mechanism. Clin Transl Immunology 2023; 12(5): e1451.
- 57. Serafini B, Rosicarelli B, Veroni C, Mazzola GA, Aloisi F. Epstein-Barr virus-specific CD8 T cells selectively infiltrate the brain in multiple sclerosis and interact locally with virus-infected cells: Clue for a virus-driven immunopathological mechanism. J Virol 2019; 93(24): 00980-19.
- 58. Hedstrom AK. Risk factors for multiple sclerosis in the context of Epstein-Barr

virus infection. Front Immunol 2023; 14: 1212676.

- Bjornevik K, Cortese M, Healy BC, Kuhle J, Mina MJ, Leng Y, et al. Longitudinal analysis reveals high prevalence of Epstein-Barr virus associated with multiple sclerosis. Science 2022; 375(6578): 296-301.
- Bar-Or A, Pender MP, Khanna R, Steinman L, Hartung HP, Maniar T, et al. Epstein-Barr virus in multiple sclerosis: Theory and emerging immunotherapies. Trends Mol Med 2020; 26(3): 296-310.
- 61. Sedighi S, Gholizadeh O, Yasamineh S, Akbarzadeh S, Amini P, Favakehi P, et al. Comprehensive investigations relationship between viral infections and multiple sclerosis pathogenesis. Curr Microbiol 2022; 80(1): 15.
- Veroni C, Aloisi F. The CD8 T Cell-Epstein-Barr Virus-B cell trialogue: A central issue in multiple sclerosis pathogenesis. Front Immunol 2021; 12: 665718.
- Kakalacheva K, Munz C, Lunemann JD. Viral triggers of multiple sclerosis. Biochim Biophys Acta 2011; 1812(2): 132-40.
- Voumvourakis KI, Fragkou PC, Kitsos DK, Foska K, Chondrogianni M, Tsiodras S. Human herpesvirus 6 infection as a trigger of multiple sclerosis: An update of recent literature. BMC Neurol 2022; 22(1): 57.
- 65. Keyvani H, Zahednasab H, Aljanabi HAA, Asadi M, Mirzaei R, Esghaei M, et al. The role of human herpesvirus-6 and inflammatory markers in the pathogenesis of multiple sclerosis. J Neuroimmunol 2020; 346: 577313.
- 66. Jeanne Billioux B, Alvarez Lafuente R, Jacobson S. HHV-6 and multiple sclerosis. Human herpesviruses HHV-6A, HHV-6B & HHV-7 2014: 123-42.
- Leibovitch EC, Jacobson S. Evidence linking HHV-6 with multiple sclerosis: An update. Curr Opin Virol 2014; 9: 127-33.
- Morris G, Maes M, Murdjeva M, Puri BK. Do human endogenous retroviruses contribute to multiple sclerosis, and if so, how? Mol Neurobiol 2019; 56(4): 2590-605.
- Donati D. Viral infections and multiple sclerosis. Drug Discov Today Dis Models 2020; 32: 27-33.
- Uzun N. Chlamydia infection's role in neurological diseases. In: Sarier M, editor. Chlamydia - secret enemy from past to present. Intechopen [Online 2023]. [cited 2023 Apr 20]; Available from: URL: https://www.intechopen.com/chapters/86 946
- Pastor Bandeira I, Silva C, Martin M, Silva GF, Parolin L, Melo L, et al. Latent tuberculosis infection in patients with multiple sclerosis [Preprints]. 2020.
- 72. Esmailkhani A, Akhi MT, Sadeghi J, Niknafs B, Zahedi BA, Farzadi L, et al. Assessing the prevalence of Staphylococcus aureus in infertile male patients in Tabriz, northwest Iran. Int J Reprod Biomed 2018; 16(7): 469-74.

- Zarghami A, Li Y, Claflin SB, van der Mei I, Taylor BV. Role of environmental factors in multiple sclerosis. Expert Rev Neurother 2021; 21(12): 1389-408.
- Correale J, Farez M. Association between parasite infection and immune responses in multiple sclerosis. Ann Neurol 2007; 61(2): 97-108.
- Karpus WJ. Cytokines and chemokines in the pathogenesis of experimental autoimmune encephalomyelitis. J Immunol 2020; 204(2): 316-26.
- Arjmandi D, Graeili Z, Mohammadi P, Arshadi M, Jafari TM, Ardekani A, et al. Chlamydia pneumonia infection and risk of multiple sclerosis: A meta-analysis. Mult Scler Relat Disord 2023; 77: 104862.
- 77. Sriram S, Stratton CW, Yao S, Tharp A, Ding L, Bannan JD, et al. Chlamydia pneumoniae infection of the central nervous system in multiple sclerosis. Ann Neurol 1999; 46(1): 6-14.
- Doran KS, Banerjee A, Disson O, Lecuit M. Concepts and mechanisms: crossing host barriers. Cold Spring Harb Perspect Med 2013; 3(7): a010090.
- 79. Kazemi S, Lashtoo Aghaee B, Soltanian AR, Mazdeh M, Taheri M, Alikhani MY. Investigation of chlamydia pneumoniae infection in patients with multiple sclerosis: A case-control study. Avicenna J Clin Microbiol Infect 2020; 7(2): 36-9.
- Woods JJ, Skelding KA, Martin KL, Aryal R, Sontag E, Johnstone DM, et al. Assessment of evidence for or against contributions of Chlamydia pneumoniae infections to Alzheimer's disease etiology. Brain Behav Immun 2020; 83: 22-32.
- Libbey JE, Cusick MF, Fujinami RS. Role of pathogens in multiple sclerosis. Int Rev Immunol 2014; 33(4): 266-83.
- Huang Y, Wang QL, Cheng DD, Xu WT, Lu NH. Adhesion and invasion of gastric mucosa epithelial cells by Helicobacter pylori. Front Cell Infect Microbiol 2016; 6: 159.
- 83. Efthymiou G, Dardiotis E, Liaskos C, Marou E, Tsimourtou V, Rigopoulou EI, et al. Immune responses against Helicobacter pylori-specific antigens differentiate relapsing remitting from secondary progressive multiple sclerosis. Sci Rep 2017; 7(1): 7929.
- Mirmosayyeb O, Barzegar M, Nehzat N, Najdaghi S, Ansari B, Shaygannejad V. Association of helicobacter pylori with multiple sclerosis: Protective or risk factor? Curr J Neurol 2020; 19(2): 59-66.
- Gavalas E, Kountouras J, Boziki M, Zavos C, Polyzos SA, Vlachaki E, et al. Relationship between Helicobacter pylori infection and multiple sclerosis. Ann Gastroenterol 2015; 28(3): 353-6.
- 86. Cook KW, Crooks J, Hussain K, O'Brien K, Braitch M, Kareem H, et al. Helicobacter pylori infection reduces disease severity in an experimental model of multiple sclerosis. Front Microbiol 2015; 6: 52.
- Kountouras J, Zavos C, Polyzos SA, Deretzi G. The gut-brain axis: Interactions between Helicobacter pylori and enteric

and central nervous systems. Ann Gastroenterol 2015; 28(4): 506.

- Whiteside SK, Snook JP, Ma Y, Sonderegger FL, Fisher C, Petersen C, et al. IL-10 Deficiency reveals a role for TLR2-dependent bystander activation of T cells in Lyme Arthritis. J Immunol 2018; 200(4): 1457-70.
- Tracy KE, Baumgarth N. Borrelia burgdorferi manipulates innate and adaptive immunity to establish persistence in rodent reservoir hosts. Front Immunol 2017; 8: 116.
- Chmielewska-Badora J, Cisak E, Dutkiewicz J. Lyme borreliosis and multiple sclerosis: any connection? A seroepidemic study. Ann Agric Environ Med 2000; 7(2): 141-3.
- Chai Q, Zhang Y, Liu CH. Mycobacterium tuberculosis: An adaptable pathogen associated with multiple human diseases. Front Cell Infect Microbiol 2018; 8: 158.
- 92. Kiazyk S, Ball TB. Latent tuberculosis infection: An overview. Can Commun Dis Rep 2017; 43(3-4): 62-6.
- Cossu D, Yokoyama K, Hattori N. Conflicting role of mycobacterium species in multiple sclerosis. Front Neurol 2017; 8: 216.
- 94. Lehmann D, Ben-Nun A. Bacterial agents protect against autoimmune disease. I. Mice pre-exposed to Bordetella pertussis or Mycobacterium tuberculosis are highly refractory to induction of experimental autoimmune encephalomyelitis. J Autoimmun 1992; 5(6): 675-90.
- 95. Salvetti M, Ristori G, Buttinelli C, Fiori P, Falcone M, Britton W, et al. The immune response to mycobacterial 70-kDa heat shock proteins frequently involves autoreactive T cells and is quantitatively disregulated in multiple sclerosis. J Neuroimmunol 1996; 65(2): 143-53.
- Liverani E, Scaioli E, Cardamone C, Dal MP, Belluzzi A. Mycobacterium avium subspecies paratuberculosis in the etiology of Crohn's disease, cause or epiphenomenon? World J Gastroenterol 2014; 20(36): 13060-70.
- 97. Cossu D, Mameli G, Galleri G, Cocco E, Masala S, Frau J, et al. Human interferon regulatory factor 5 homologous epitopes of Epstein-Barr virus and Mycobacterium avium subsp. paratuberculosis induce a specific humoral and cellular immune response in multiple sclerosis patients. Mult Scler 2015; 21(8): 984-95.
- Mameli G, Cocco E, Frau J, Marrosu MG, Sechi LA. Epstein Barr Virus and Mycobacterium avium subsp. Paratuberculosis peptides are recognized in sera and cerebrospinal fluid of MS patients. Sci Rep 2016; 6: 22401.
- Arsenault RJ, Li Y, Maattanen P, Scruten E, Doig K, Potter A, et al. Altered Toll-like receptor 9 signaling in Mycobacterium avium subsp. paratuberculosis-infected bovine monocytes reveals potential therapeutic targets. Infect Immun 2013; 81(1): 226-37.
- 100.Cossu D, Masala S, Sechi LA. A Sardinian map for multiple sclerosis. Future

Microbiol 2013; 8(2): 223-32.

- 101.Ramos M, Pisa D, Molina S, Rabano A, Juarranz A, Carrasco L. Fungal infection in patients with multiple sclerosis. The Open Mycology Journal 2008; 2(1): 22-8.
- 102.Mangalam AK. Fungal microbiome and multiple sclerosis: The not-so-new kid on the block. EBioMedicine 2021; 72: 103621.
- 103.Tanasescu R, Tench CR, Constantinescu CS, Telford G, Singh S, Frakich N, et al. Hookworm treatment for relapsing multiple sclerosis: A randomized doubleblinded placebo-controlled trial. JAMA Neurol 2020; 77(9): 1089-98.
- 104. Yordanova IA, Ebner F, Schulz AR, Steinfelder S, Rosche B, Bolze A, et al. The worm-specific immune response in multiple sclerosis patients receiving controlled Trichuris suis Ova immunotherapy. Life (Basel) 2021; 11(2): 101.
- 105.Scotto R, Reia A, Buonomo AR, Moccia M, Viceconte G, Pisano E, et al. Risk of invasive fungal infections among patients treated with disease modifying treatments for multiple sclerosis: A comprehensive review. Expert Opin Drug Saf 2021; 20(8): 925-36.
- 106.Benito-Leon J, Laurence M. The role of fungi in the etiology of multiple sclerosis. Front Neurol 2017; 8: 535.
- 107.Goldszmid RS, Caspar P, Rivollier A, White S, Dzutsev A, Hieny S, et al. NK cell-derived interferon-gamma orchestrates cellular dynamics and the differentiation of monocytes into dendritic cells at the site of infection. Immunity 2012; 36(6): 1047-59.
- 108.Keighobadi M, Alokandeh ND, Baghbanian SM, Karimi N. The role of Toxoplasma gondii in multiple sclerosis: A matched case-control study. Neurology Asia 2021; 26(2): 333-9.
- 109.Nicoletti A, Cicero CE, Giuliano L, Todaro V, Lo Fermo S, Chisari C, et al. Toxoplasma gondii and multiple sclerosis: A population-based case-control study. Sci Rep 2020; 10(1): 18855.
- 110.Rahnama M, Asgari Q, Petramfar P, Tasa D, Hemati V, et al. The role of Toxoplasma gondii infection among multiple sclerosis patient compared to ordinary people in south of Iran: A Case-Control Study. Mod Care J 2020; 17(3): e105090.
- 111.Marignier R, Hacohen Y, Cobo-Calvo A, Probstel AK, Aktas O, Alexopoulos H, et al. Myelin-oligodendrocyte glycoprotein antibody-associated disease. Lancet Neurol 2021; 20(9): 762-72.
- 112.Sotgiu S, Sannella AR, Conti B, Arru G, Fois ML, Sanna A, et al. Multiple sclerosis and anti-Plasmodium falciparum innate immune response. J Neuroimmunol 2007; 185(1-2): 201-7.
- 113.Belbasis L, Bellou V, Evangelou E, Tzoulaki I. Environmental factors and risk of multiple sclerosis: Findings from metaanalyses and Mendelian randomization studies. Mult Scler 2020; 26(4): 397-404.
- 114.Omerhoca S, Akkas SY, Icen NK.

Multiple sclerosis: Diagnosis and differential diagnosis. Noro Psikiyatr Ars 2018; 55(Suppl 1): S1-S9.

- 115. Asouri M, Sahraian MA, Karimpoor M, Fattahi S, Motamed N, Doosti R, et al. Molecular detection of epstein-barr virus, human herpes virus 6, cytomegalovirus, and hepatitis B virus in patients with multiple sclerosis. Middle East J Dig Dis 2020; 12(3): 171-7.
- 116.Ruprecht K. The role of Epstein-Barr virus in the etiology of multiple sclerosis: A current review. Expert Rev Clin Immunol 2020; 16(12): 1143-57.
- 117.Khalesi Z, Tamrchi V, Razizadeh MH, Letafati A, Moradi P, Habibi A, et al. Association between human herpesviruses and multiple sclerosis: A systematic review and meta-analysis. Microb Pathog 2023; 177: 106031.
- 118.Jakhmola S, Upadhyay A, Jain K, Mishra A, Jha HC. Herpesviruses and the hidden links to Multiple Sclerosis neuropathology. J Neuroimmunol 2021; 358: 577636.
- 119.Xu Y, Hiyoshi A, Smith KA, Piehl F, Olsson T, Fall K, et al. Association of infectious mononucleosis in childhood and adolescence with risk for a subsequent multiple sclerosis diagnosis among siblings. JAMA Netw Open 2021; 4(10): e2124932.
- 120.Perlejewski K, Bukowska-Osko I, Rydzanicz M, Dzieciatkowski T, Zakrzewska-Pniewska B, Podlecka-Pietowska A, et al. Search for viral agents in cerebrospinal fluid in patients with multiple sclerosis using real-time PCR and metagenomics. PLoS One 2020; 15(10): e0240601.
- 121.Pagani CF, Curbelo MC, Vazquez G, Sedeno L, Steinberg J, Carra A, et al. Application of the 2017 McDonald criteria for the diagnosis of multiple sclerosis after a first demyelinating event in patients from Argentina. Mult Scler Relat Disord 2020; 41: 102043.
- 122.Tan IL, McArthur JC, Clifford DB, Major EO, Nath A. Immune reconstitution inflammatory syndrome in natalizumabassociated PML. Neurology 2011; 77(11): 1061-7.
- 123.Messe A, Caplain S, Paradot G, Garrigue D, Mineo JF, Soto AG, et al. Diffusion tensor imaging and white matter lesions at the subacute stage in mild traumatic brain injury with persistent neurobehavioral impairment. Hum Brain Mapp 2011; 32(6): 999-1011.
- 124.Shah R, Bag AK, Chapman PR, Cure JK. Imaging manifestations of progressive multifocal leukoencephalopathy. Clin Radiol 2010; 65(6): 431-9.
- 125.Lycke J. Trials of antivirals in the treatment of multiple sclerosis. Acta Neurol Scand 2017; 136 Suppl 201: 45-8.
- 126. Yang JH, Rempe T, Whitmire N, Dunn-Pirio A, Graves JS. Therapeutic advances in multiple sclerosis. Front Neurol 2022; 13: 824926.
- 127.Giovannoni G. Personalized medicine in multiple sclerosis. Neurodegener Dis Manag 2017; 7(6s): 13-7.

- 128.Hosseinzadeh A, Iranpour S, Adineh HA, Aliyari R. Antibiotic use and multiple sclerosis: A systematic review and meta-analysis. Mult Scler Relat Disord 2023; 75: 104765.
- 129.Mendes A, Sa MJ. Classical immunomodulatory therapy in multiple sclerosis: How it acts, how it works. Arq Neuropsiquiatr 2011; 69(3): 536-43.
- 130.Rahmani M, Negro Alvarez SE, Hernandez EB. The potential use of tetracyclines in neurodegenerative diseases and the role of nano-based drug delivery systems. Eur J Pharm Sci 2022; 175: 106237.
- 131.Wang D, Lu Z, Hu L, Zhang Y, Hu X. Macrolide antibiotics aggravate experimental autoimmune encephalomyelitis and inhibit inducible nitric oxide synthase. Immunol Invest 2009; 38(7): 602-12.
- 132.Dumitrescu L, Papathanasiou A, Coclitu C, Constantinescu CS, Popescu BO, Tanasescu R. Beta interferons as immunotherapy in multiple sclerosis: A new outlook on a classic drug during the COVID-19 pandemic. QJM 2021; 114(10): 691-7.
- 133.Bourque J, Hawiger D. Current and future immunotherapies for multiple sclerosis.

Mo Med 2021; 118(4): 334-9.

- 134.Freedman MS, Selchen D, Prat A, Giacomini PS. Managing multiple sclerosis: Treatment initiation, modification, and sequencing. Can J Neurol Sci 2018; 45(5): 489-503.
- 135.Wingerchuk DM, Carter JL. Multiple sclerosis: current and emerging diseasemodifying therapies and treatment strategies. Mayo Clin Proc 2014; 89(2): 225-40.
- 136. Greenberg BM, Calabresi PA. Future research directions in multiple sclerosis therapies. Semin Neurol 2008; 28(1): 121-7.